

Challenges in Genetic Interpretation of Hereditary Kidney Diseases

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RAPIDS
Renal Alliance for Precision Diagnosis in Singapore

National University Hospital
National University Centre for Genomic Medicine



DISCLOSURES

- No conflicts of interest

A Typical Kidney Biopsy Report

Most nephrologists are comfortable interpreting this

ADDENDUM/AMENDMENT:

ADDENDUM: HISTOPATHOLOGY REPORT 25:PR919

Toluidine blue stained semithin sections

The EM sample contains 5 glomeruli.

The glomeruli show increase in mesangial matrix. No overt tubular atrophy is seen; focal interstitial fibrosis is noted.

Ultrastructural study

There is approximately 10% effacement of podocyte foot processes. The glomerular basement membranes are generally of normal thickness, though there are segments which are slightly thinner (135nm) and thicker (599nm). Increased mesangial matrix is noted. Mesangial and focal para-mesangial / subendothelial electron dense deposits are identified. No tubuloreticular structures are noted.

Conclusion:

The electron microscopy findings are supportive of the original diagnoses.

NOTE: Please see below for original report.

Pathologist :



DR CHNG TZE WEI

13/11/25 1459 hrs

DIAGNOSIS:

<AP-AII>

Singapore General Hospital Pte Ltd
Outram Road, Singapore 169608
www.sgh.com.sg
Reg No 1987039072

Renal biopsy:
Overall, features are in keeping with IgA nephropathy.

Comment:
Pending ultrastructural studies.

CLINICAL HISTORY:

12 year old male

- Subnephrotic proteinuria and haematuria for more than 1 year
- No family history or extra-renal conditions
- Normal eGFR
- Genetic test: VUS in COL4A3
- DDx: IgAN, Alport's syndrome

GROSS DESCRIPTION:

(A) The specimen is received in Bouin's fixative, labelled with patient's data and designated "renal biopsy". It consists of 1 core of tissue measuring 1.5cm in length.
(A1; no reserve)

(B) Fresh tissue received for immunofluorescence. It consists of a 1 core of tissue measuring 0.25cm in length.
(B1; no reserve)

(C) Tissue received in glutaraldehyde for electron microscopy. It consists of 2 cores tissue measuring 0.1cm in length.
(C1; no reserve)

MICROSCOPIC DESCRIPTION:

Specimen type: Native kidney.

Light microscopy:

Sections show 1 core of renal cortico-medullary tissue containing 5 glomeruli, of which none are globally or almost globally sclerosed (%).

GLOMERULI

There is mild mesangial matrix expansion with without overt mesangial hypercellularity. No endocapillary hypercellularity is seen. No areas of segmental sclerosis or crescents are seen. Special stains do not highlight membranous spikes/vacuoles or obvious deposits.

<AP-AII>

TUBULES
No overt tubular atrophy is noted.

INTERSTITIUM
No overt inflammation or fibrosis is seen.

VESSELS
No atherosclerosis or arteriolosclerosis are seen.

Oxford Classification for IgA nephropathy (2016 update): M0 E0 S0 T0 C0

IMMUNOFLUORESCENCE

No. of gloms present: 12

IgG	: Negative.
IgA	: 3+ mesangial.
IgM	: 1+ mesangial.
C3	: 2+ to 3+ mesangial.
C1q	: Negative.
Albumin	: Negative.
Lambda	: 3+ mesangium.
Kappa	: 1+ mesangium.
C4	: Negative.
Fibrinogen	: Negative.

Collagen (a5+a2) = Glomerular basement membranes and Bowman's capsule + scattered (distal) tubules + (slightly weak).

Pathologist :

DR CHNG TZE WEI
28/08/25 1701 hrs

<AP-AII>

Singapore General Hospital Pte Ltd
Outram Road, Singapore 169608
www.sgh.com.sg
Reg No 1987039072

A Typical Genetic Report

Most nephrologists are NOT comfortable interpreting this

Final Report: 3/1/2024

[COL4A4 Additional Information](#)

GENE INFORMATION:

All content hereafter is supplemental information to the preceding report.

[Variant\(s\) of Uncertain Significance](#)

SUMMARY

POSITIVE: Pathogenic & Likely Pathogenic

RESULTS

Gene	Inheritance	Alteration	Proband
COL4A4 (NM_000092)	Autosomal dominant, Autosomal recessive	Variant, Likely Pathogenic: c.2752G>A (p.Gly918Arg) Pathogenic Mutation	Heterozygous

INTERPRETATION

- This individual is heterozygous for the c.2752G>A (p.Gly918Arg) likely pathogenic mutation in the *COL4A4* gene.
- This result is consistent with a diagnosis of *COL4A4*-related Alport syndrome.
- Familial testing would be necessary to determine if these alterations are on the same or different chromosomes (in *cis* or *trans*).
- The expression and severity of disease for this individual cannot be predicted.
- Genetic testing for pathogenic and/or likely pathogenic variants in family members can be helpful in identifying at-risk individuals.
- Genetic counseling is a recommended option for all individuals undergoing genetic testing.

VARIANT DETAILS:

- The c.2752G>A (p.Gly918Arg) alteration is located in exon 31 (coding exon 30) of the *COL4A4* gene. This alteration results from a G to A substitution at nucleotide position 2752, causing the glycine (G) at amino acid position 918 to be replaced by arginine (R).
- This variant was reported in multiple individuals.

REFERENCES:

- Verduciello P, et al. (2022) *Front Med* 9:836283. PMID:35419377
- Fallerini C, et al. (2021) *Front Med* 9:734412. PMID:343675912

VARIANT DETAILS:

- The c.3967C>T (p.Q132*) alteration, located in exon 41 (coding exon 40) of the *COL4A4* gene, consists of a C to T substitution at nucleotide position 3967. This changes the amino acid from a glutamine (Q) to a stop codon at amino acid position 1323. This alteration is expected to result in loss of function by premature protein truncation or nonsense-mediated mRNA decay.
- This variant has been reported in the heterozygous state, and in conjunction with another alteration in *COL4A4*, in individuals with clinical features of Alport syndrome.

REFERENCES:

- Xie J, et al. (2014) *J Mol Cell Biol* 6(6):498-505. PMID:25596306
- Zhang Y, et al. (2021) *Pediatr Nephrol* 36(9):2719-2730. PMID:33772369
- Zhou L, et al. (2023) *J Nephrol* 36(5):1415-1423. PMID:37079554

Genes Analyzed

(84 total): *ACTN4, ADAMTS13, AGXT, AMN, ANLN, APOL1, ARHGDIA, C3, CD2AP, CD46, CFB, CFH, CFHRE, CFI, CLDN5, COL4A3, COL4A4, COL4A5, COQ2, COO6, COQ8B, CRB2, CTNS, CUBN, DGKE, DHTK101, DLC1, EMP2, FAN1, FAT1, FEN1, GLA, GRIPR, INF2, ITGB4, KANK1, KANK2, KANK4, LAGE3, LAMAS, LAMB2, LMX1B, LYZ, MAFB, MAGI2, MYH6, MYO1E, NPHS1, NPHS2, NUP107, NUP133, NUP160, NUP203, NUP85, NUP93, NXF5, OCRL, OSGP1, PAX6, POSS1, PDS2, PLC1, PDXK, PTEN, REN, SCARB2, SEC61AT1, SGPL1, SMARCAL1, TSC1DB8, THBD, TPSRK, TRIM33, TRPC6, TTC21B, TTR, UMOD, WDR4, WDR73, WT1 and XPO5*

and is available for download through AmbryPort or can be e-mailed by request.

Id not achieve 100% coverage at 10X for all nucleotides in the coding regions:

PODXL (94.28%), **TRPC6** (97.32%)

covered at $\geq 10X$

Genetic reports will become as common as biopsy reports

Laboratory Director: Chia-Ling Gau, PhD, DABMGG, CLIA# 05D0981414
Toll Free 866.262.7943 | Ph 949.900.5500 | Fx 949.900.5501 | [www.ambrygen.com](#) | 7 Argonaut, Aliso Viejo, CA 92656

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Outline: Interpreting genetic reports

- Basic 101: What nephrologists should know
- Understanding the limitations and caveats

Assessing the adequacy of the biopsy

DIAGNOSIS:

RENAL BIOPSY:

- DIFFUSE PROLIFERATIVE GLOMERULONEPHRITIS WITH CELLULAR CRESCENTS CONSISTENT WITH IgA DISEASE.

GROSS DESCRIPTION:

The specimen is received in Bouin's fixative and labelled with patient's data. It consists of a core of tissue measuring 1.0cm in length. (A1, no reserve)

MICROSCOPIC DESCRIPTION:

Sections show one core of renal cortical tissue with 17 glomeruli.

Glomeruli:

One glomerulus is globally sclerosed. Most glomeruli show increase in mesangial matrix and increase in mesangial cellularity. Cellular crescent are seen in 4 glomeruli (24%). Endocapillary proliferation is seen in one glomerulus. Capillary loops are patent and no double contours are seen. Occasional circulating leucocytes are seen in the capillary lumen. Karyorrhectic particles and fibrinoid necrosis are not seen. Hyaline thrombi are not identified. Masson-silver stains do not show fuchsinophilic subepithelial deposits or argyrophilic basement membrane spikes.

Tubules:

No tubular changes are seen. No acute tubular necrosis is seen.

Interstitium:

Mild

	Immunofluorescence:	Six glomeruli are present, one is sclerotic.
IgG	:	Negative.
IgA	:	3+ mesangium and segmentally along the glomerular capillary walls.
IgM	:	1+ to 2+ mesangium segmentally.
C3	:	1+ to 2+ diffuse granular staining of mesangium.
C1q	:	Trace mesangium segmentally.
Albumin	:	Negative.
L	:	2+ to 3+ diffuse granular staining of mesangium and some tubular casts.
K	:	2+ to 3+ diffuse granular staining of mesangium and some tubular casts.
C4	:	Negative.
Fibrin	:	Negative.

1) Tests:

Light microscopy

Immunofluorescence

Electron microscopy

2) Clinical history

3) Sample amount:

Number of glomeruli

Cortex, corticomedullary, medulla

Assessing the adequacy of the genetic analysis



Final Report: 3/1/2024

Patient Name: [REDACTED]
MRN #: NPM Accession #: 24-096784

3) Exome/ Genome: Clinical history submitted to laboratory

ExomeNext®-Select: Analysis of Selected Genes

SUMMARY

POSITIVE: Pathogenic & Likely Pathogenic Alterations Detected

RESULTS

Gene	Inheritance	Proband
COL4A4 (NM_000092)	Autosomal	G>A (p.G918R) Heterozygous
		T (p.Q1323*) Heterozygous

INTERPRETATION

- This individual is heterozygous for a pathogenic mutation in the COL4A4 gene.
- This result is consistent with the clinical presentation.
- Familial testing would be recommended.
- The expression and severity of the disease are variable.
- Genetic testing for pathogenic variants is available.
- Genetic counseling is available.

1) What kind of test?
Gene panel
Targeted exome
Exome
Genome

This individual was also found to have a heterozygous mutation in the ADAMTS13 gene, which may or may not contribute to this individual's clinical history. Refer to the supplemental pages for additional information on this variant. No additional pathogenic mutations, variants of uncertain significance, or gross deletions or duplications were detected.

All content hereafter is supplemental information to the preceding report.

Variant(s) of Uncertain Significance

Gene (RefSeq ID)	Associated Condition(s) (Inheritance) ^{1,2}	Alteration	Population Frequency ³	In Silico ⁴	Notes/ References	Proband
ADAMTS13 (NM_139025)	ADAMTS13-related thrombotic thrombocytopenic purpura (AR)	c.215_220delAGAGGC (p.Q72_R73del)	0.02%	N/A	N/A	Heterozygous

Is the suspected gene included?
Which genes did not achieve adequate coverage 10-30x?

Report

- COL4A4
- ADAMTS13
- Gao Y, et al. (2022) *Front Genet* 13:1054491. PMID:36699462
- Isaranuwatchai S, et al. (2023) *Sci Rep* 13(1):805. PMID:36646731
- Lee JM, et al. (2019) *J Clin Med* 8(2):178. PMID:30717457
- Longo I, et al. (2006) *Nephrol. Dial. Transplant.* 21(3):665-71. PMID:16338941
- Lu L, et al. (2022) *Clin Genet* 101(5):541-551. PMID:35403022
- Papazachariou L, et al. (2019) *Am J Med Genet A* 176(1):10-16. PMID:30717457
- Savage J, et al. (2022) *Clin Genet* 107(2):145-152. PMID:35403022
- Storey H, et al. (2013) *J. Am Geriatr Soc* 61(10):1033-1038. PMID:24103022
- Xie J, et al. (2014) *J Mol Cell Cardiol* 69:10-16. PMID:24103022
- Zhang Y, et al. (2021) *Pediatr Nephrol* 36(10):2671-2678. PMID:34088022
- Zhou L, et al. (2023) *J Nephrol* 36(10):2671-2678. PMID:34088022

2) List of genes analyzed
Metrix and Coverage

Genes Analyzed

(94 total): *ACTN4, ADAMTS13, AGXT, AMN, ANLN, APO1, ARHGAP24, ARHGDI1, C3, CD2AP, CD46, CFB, CFHR, CFI, CLCN5, COL4A3, COL4A4, COL4A5, COQ2, COQ6, COQ8B, CRB2, CTNS, CUBN, DGKE, DHTKD1, DLC1, EMP2, FAN1, FAT1, FN1, GLA, GRHPR, INF2, ITGA3, ITGB4, KANK1, KANK2, KANK4, LAGE3, LAMA5, LAMB2, LMX1B, LYZ, MAFB, MAGI2, MYH9, MYO1E, NPHS1, NPHS2, NUP107, NUP133, NUP160, NUP205, NUP85, NUP93, NXF5, OCLR, OSGER, PAX2, PDSS1, PDSS2, PLCE1, PODXL, PTPRO, REN, SCARB2, SEC61A1, SGPL1, SMARCA1, TBC1D8B, THBD, TP53RK, TPRKB, TRIM8, TRPC6, TTC21B, TTR, UMOD, WDR4, WDR73, WTI and XPO5*

Metrics and Coverage

Complete coverage data for this proband is available for download through AmbryPort or can be e-mailed by request.

The following genes (coverage)* did not achieve 100% coverage at 10X for all nucleotides in the coding regions:

NXF5 (94.35%), PDSS1 (97.36%), PODXL (94.28%), TRPC6 (97.32%)

*percentage of the coding region covered at ≥10X

Variant interpretation: What nephrologists need to know

```
@ERR000589.41 EAS139_45:5:1:2:111/1
CTTCCTCCCTGTTCTGGCCCCACCATTCCAGGAAACATCTTGTCA
+
3IIIIIIIIII>IIIIFF9BG08E00I%IG+&?(4)%00646.C1#&(
@ERR000589.42 EAS139_45:5:1:2:1293/1
AGTTGTTAAAATCCAAGCCAATTAAGATAGTCTTATCTTTAAAAGAAAT
+
IIIIIGII.AIIII=?I9G-/II=+I=4?761BA2C9I+5A711+&>1$/I
```

6 billion nucleotides

Patient's genetic sequence

>5 million variants per person

compare

Reference sequence

GRCh37 = hg19

GRCh38 = hg38
(newer and more accurate)

Telomere-to-telomere
(T2T-CHM13)

2 disease-causing variants (high impact)

Carrier for at least 2 variants in AR diseases



Genetic variants: 5 categories

American College of Medical Genetics and Genomics (ACMG)

Association for Molecular Pathology (AMP)

Probability of
causing disease



<1%	Benign	Class 1
1-10%	Likely benign	Class 2
10-90%	Variant of unknown significance (VUS)	Class 3 No clinical impact
90-99%	Likely pathogenic	Class 4
>99%	Pathogenic	Class 5

Genetic variant X

Is this variant pathogenic?

Criteria to assign pathogenicity

(Very) Strong

BA1

Moderate

BS1 BS2 BS3 BS4

Supporting

BP1 BP2 BP3 BP4 BP5 BP6 BP7

PVS1 PS1 PS2 PS3 PS4

PM1 PM2 PM3 PM4 PM5 PM6

PP1 PP2 PP3 PP4 PP5

ACMG 2015 guidelines

PATHOGENIC

1 very strong + 1 mod+ 1 supporting
1 strong + 1 mod+ 4 supp

LIKELY PATHOGENIC

1 very strong + 1 mod
1 mod + 4 supp

Benign

Likely benign

Variant of
unknown
significance

Likely
pathogenic

Pathogenic

Benign
2 Strong

Likely Benign
1 Strong + 1 Supporting

Genetic variant X

Is this variant pathogenic?

Criteria to assign pathogenicity

(Very) Strong

BA1

Moderate

BS1 BS2 BS3 BS4

Supporting

BP1 BP2 BP3 BP4 BP5 BP6 BP7

PVS1

PS1

PS2

PS3

PS4

PM1

PM2

PM3

PM4

PM5

PM6

PP1

PP2

PP3

PP4

PP5

Bayesian framework
of scoring variants

≤ -7

Benign

Likely
pathogenic

≥ 10

Likely benign

-1 to -6

Variant of
unknown
significance

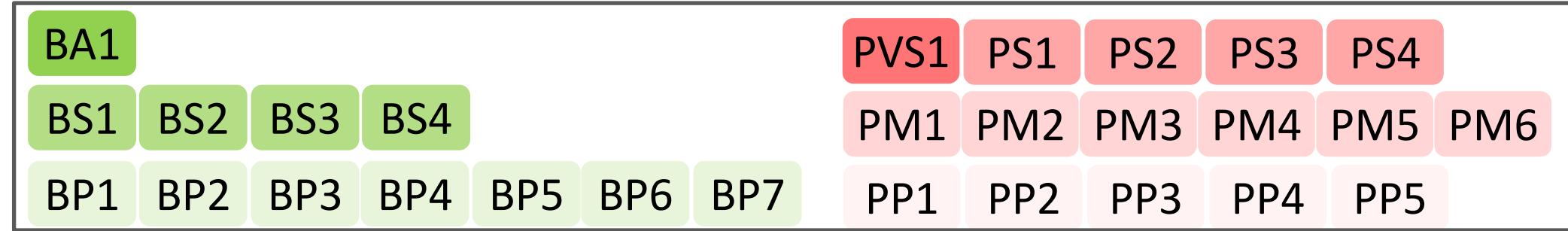
0-5

Pathogenic

6-9

Strength	Pathogenic	Benign
Indeterminate	0	0
Supporting	1	-1
Moderate	2	-2
Strong	4	-4
Very strong	8	-8

Is the variant causing the disease?



Laboratory

Population frequency

In silico tools

Nature of variant

Hot spot for mutations

Functional studies

Previous reported cases

Report:
VUS

Clinician

Gene-phenotype

Mode of inheritance

Genomic

Phenotype

specificity

Family

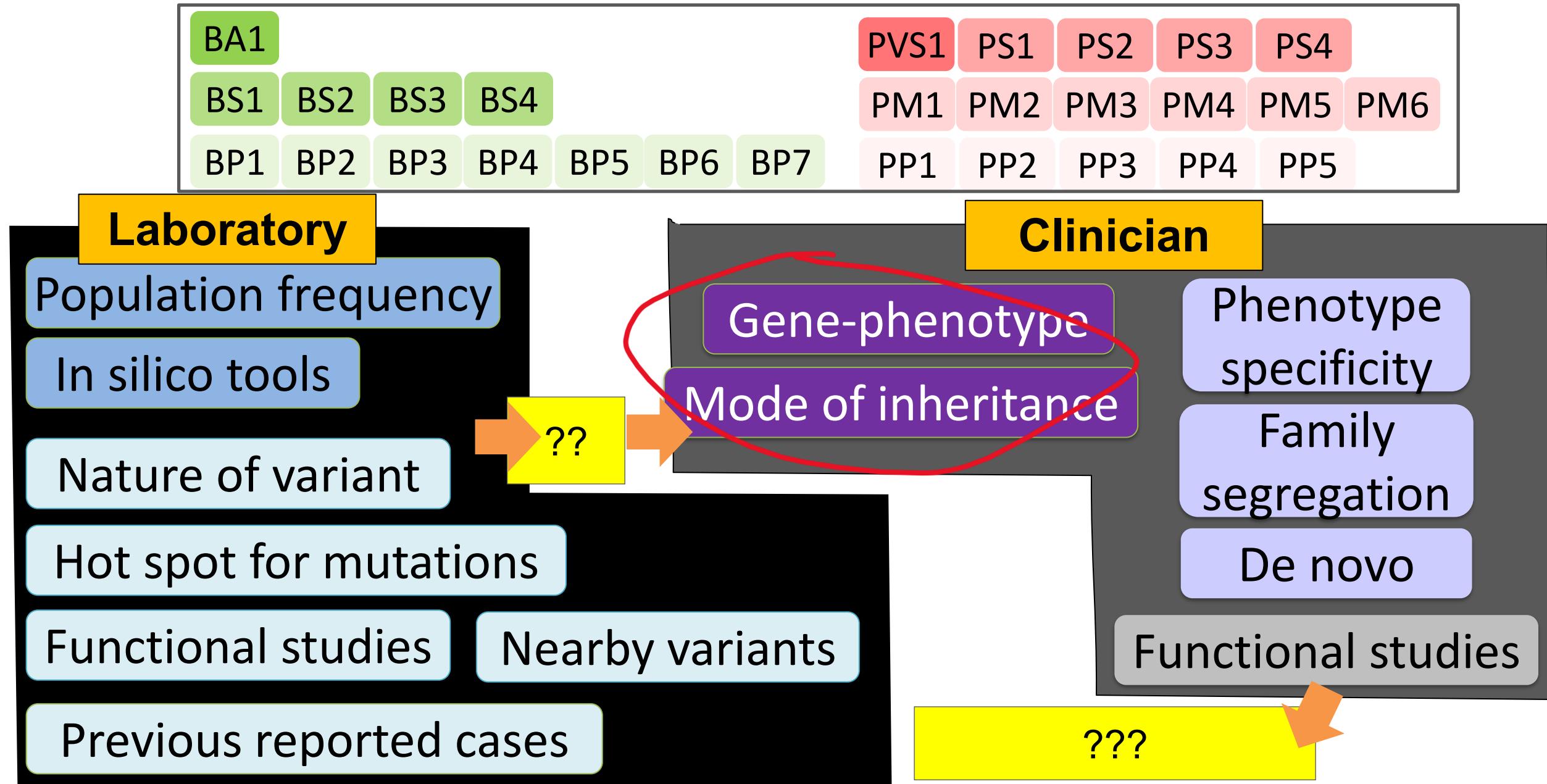
segregation

De novo

Functional studies

Likely pathogenic

Is the variant causing the disease?



Do these variants explaining the disease of the patient?

- 15 years old, Chinese boy
- Persistent isolated haematuria with no albuminuria
- Mother has microscopic haematuria

Variant(s) of Uncertain Significance

Gene (RefSeq ID)	Associated Condition(s) (Inheritance) ^{1,2}	Alteration	Population Frequency ³	In Silico ⁴	Notes/References	Proband
COL4A4 (NM_000092)	COL4A4-related Alport syndrome (AD, AR)	c.1805G>A (p.G602E) ⁵	N/A	Deleterious	Isaranuwatchai, 2023	Heterozygous
FAT1 (NM_05245)	FAT1-related nephrotic syndrome (AR)	c.13484G>A (p.P4495L)	0.11%	Tolerated	N/A	Heterozygous

Is FAT1 consistent with the phenotype?

No

Is the mode of inheritance and zygosity consistent?

No

Do these variants explaining the disease of the patient?

- 15 years old, Chinese boy
- Persistent isolated haematuria with no albuminuria
- Mother has microscopic haematuria

Variant(s) of Uncertain Significance

Gene (RefSeq ID)	Associated Condition(s) (Inheritance) ^{1,2}	Alteration	Population Frequency ³	In Silico ⁴	Notes/References	Proband
<i>COL4A4</i> (NM_000092)	<i>COL4A4</i> -related Alport syndrome (AD, AR)	c.1805G>A (p.G602E) ⁵	N/A	Deleterious	Isaranuwatchai, 2023	Heterozygous
<i>FAT1</i> (NM_05245)	FAT1-related nephrotic syndrome (AR)	c.13484G>A (p.P4495L)	0.11%	Tolerated	N/A	Heterozygous

Is *COL4A4* consistent with the phenotype?

Yes

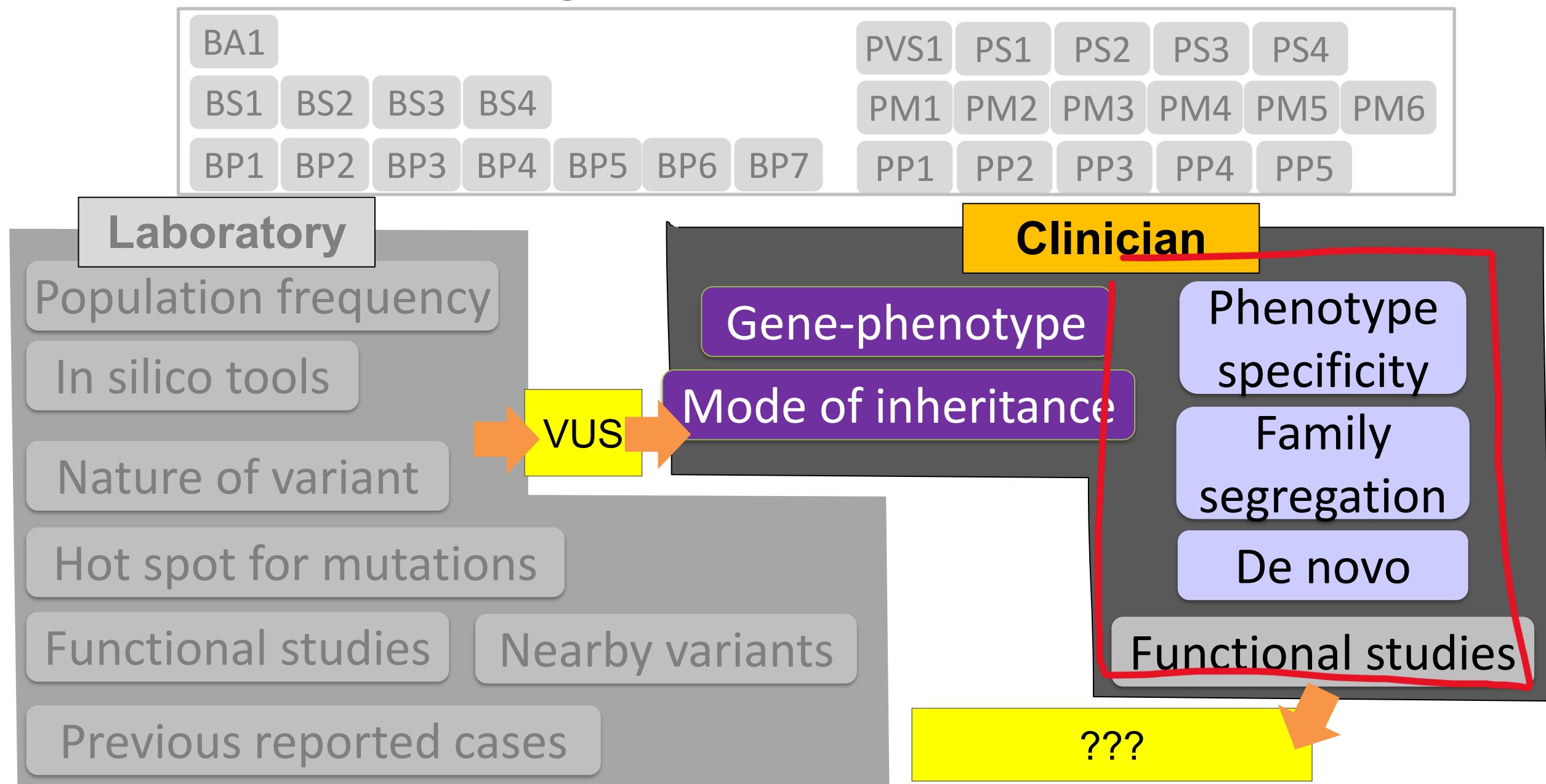
Is the mode of inheritance and zygosity consistent?

Yes

Can this *COL4A4* variant of uncertain significance be upgraded?

Maybe

Is the variant causing the disease?



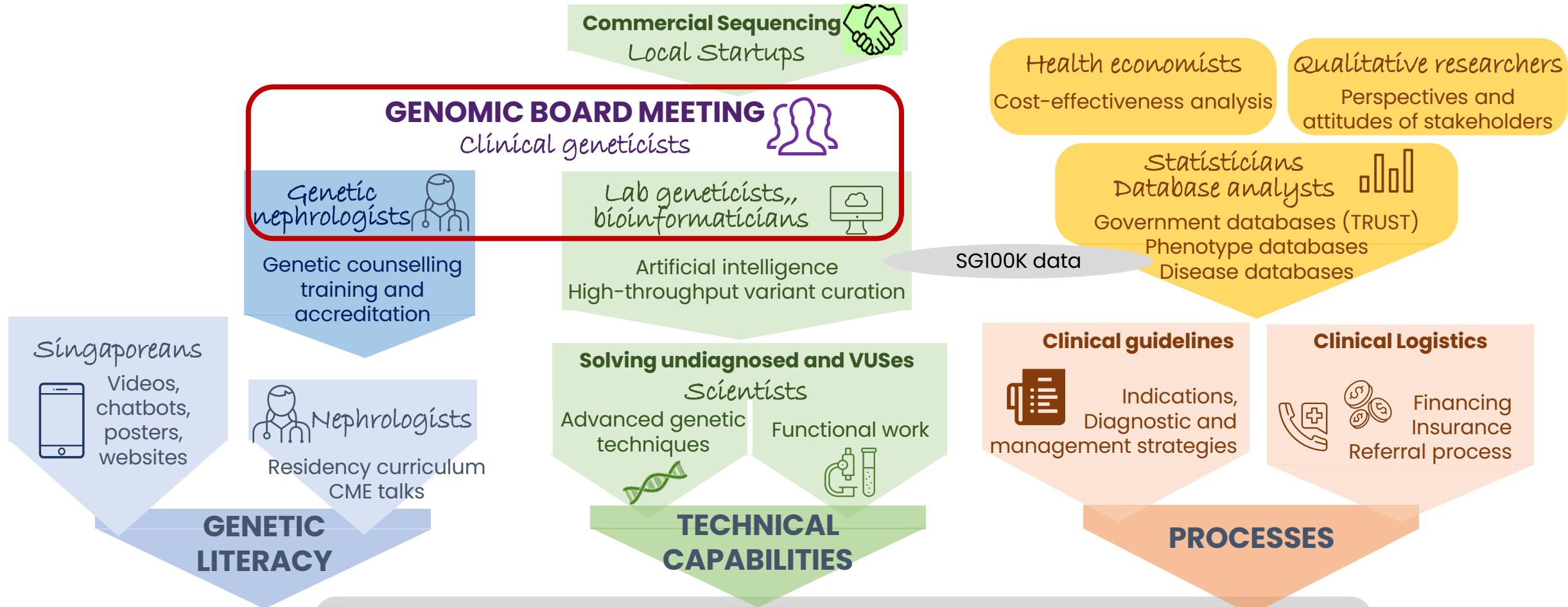
High burden of Variants of Uncertain Significance in Asia

Asian genomes represent

**6.6% of Genome Aggregation Database
(gnomAD): Lack of population-specific
allele frequency data**

**3% of genomic research studies: Lack of
functional or clinical evidence for many
variants**





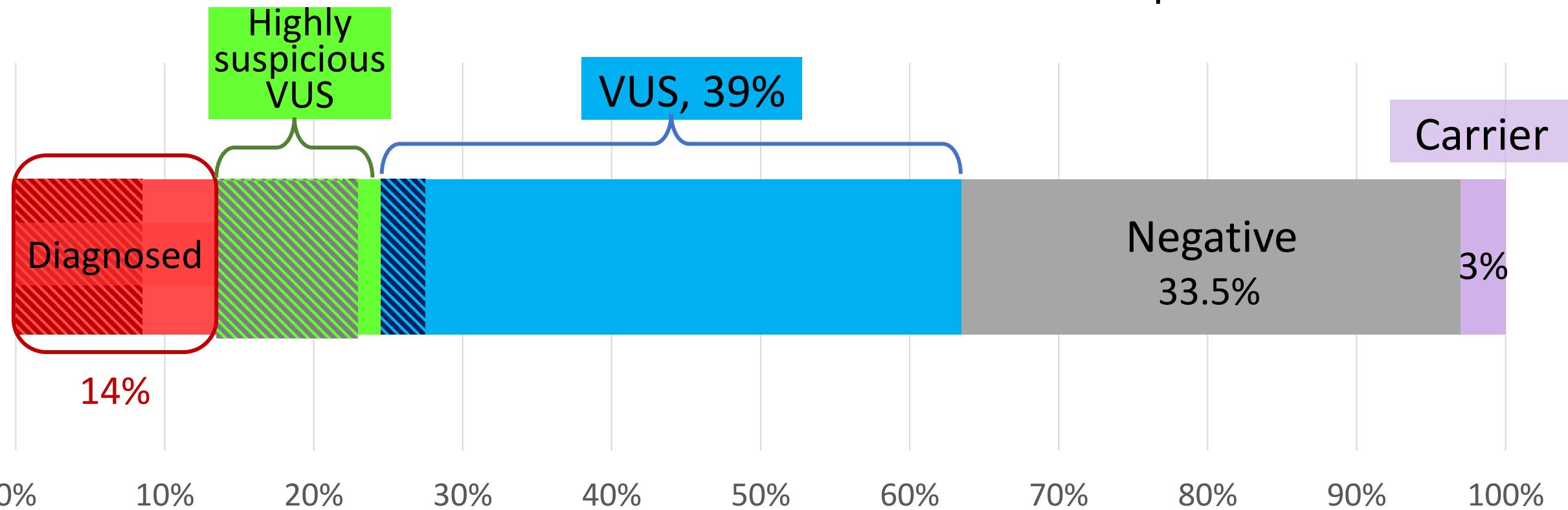
Genetic testing in nephrology clinics

Sustainable, cost-effective, evidence-based, locally-contextualised



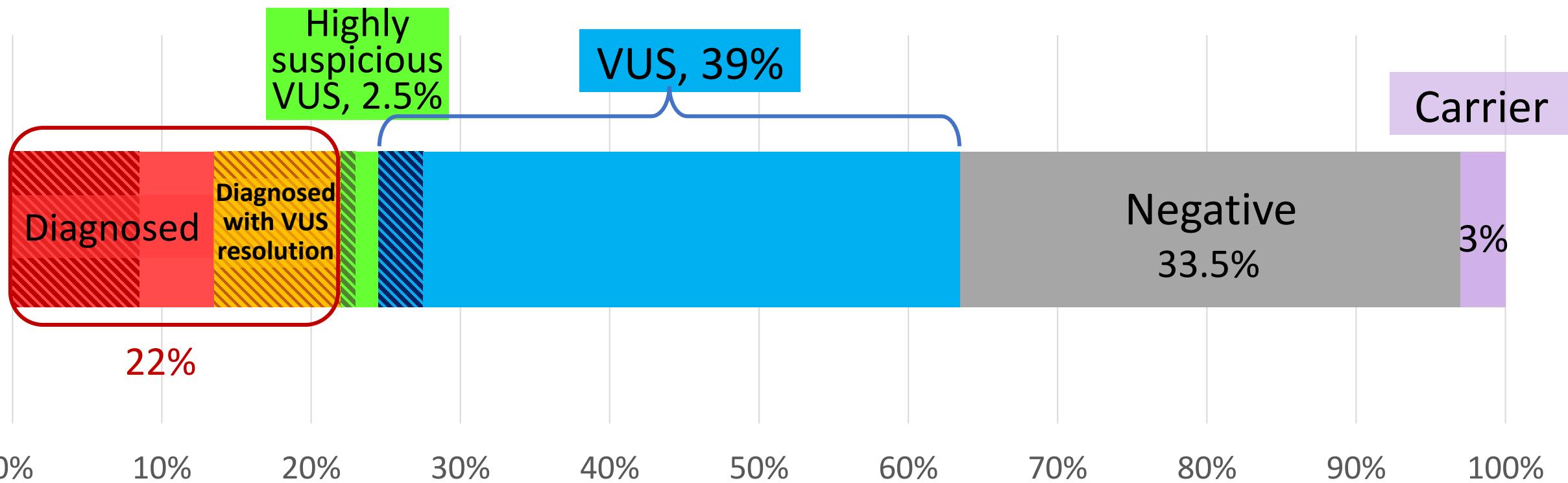
What have we found in RAPIDS?

n = 200 index patients
■ Alport



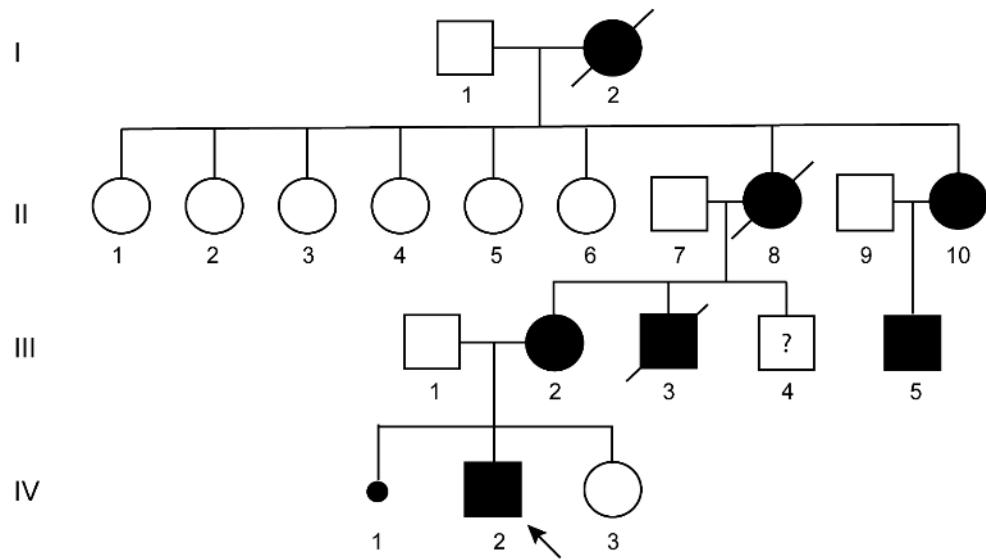
What have we found in RAPIDS?

n = 200 index patients
■ Alport

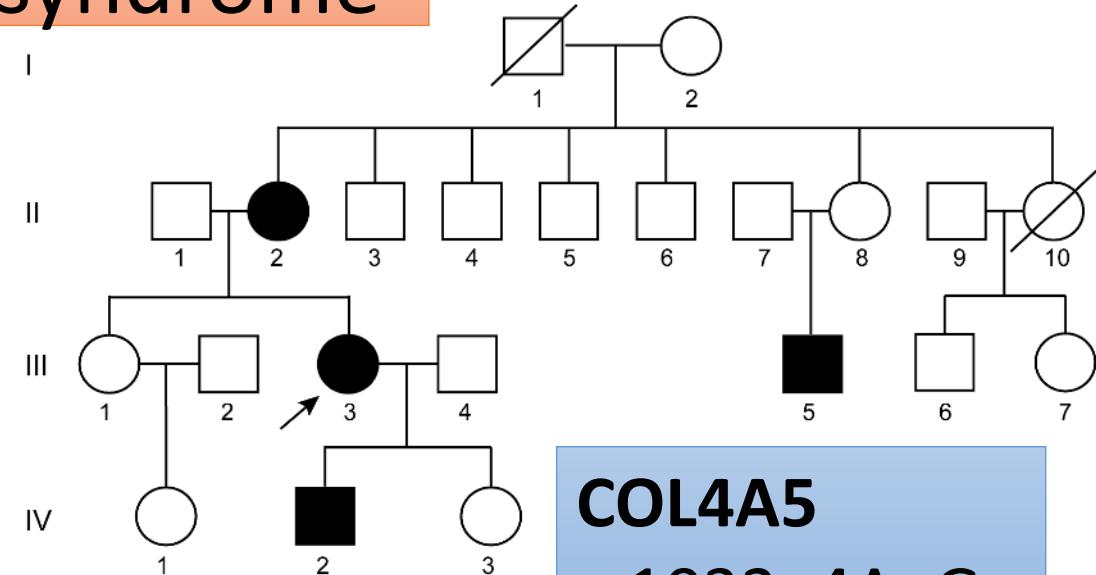


Functional studies to resolve variants of uncertain significance

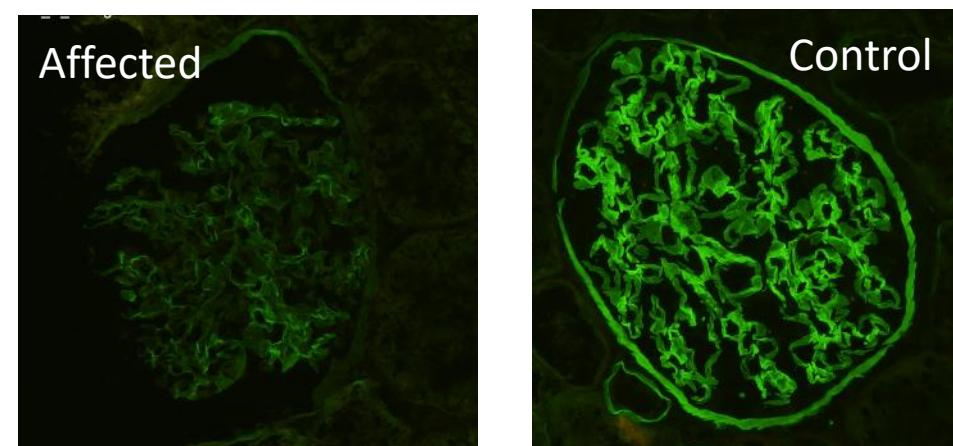
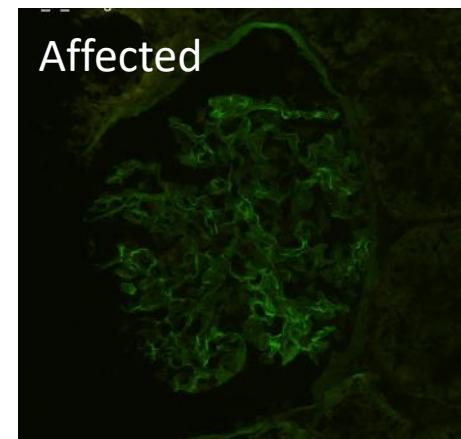
X-linked Alport syndrome



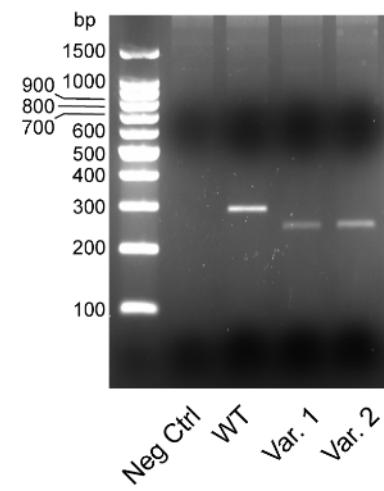
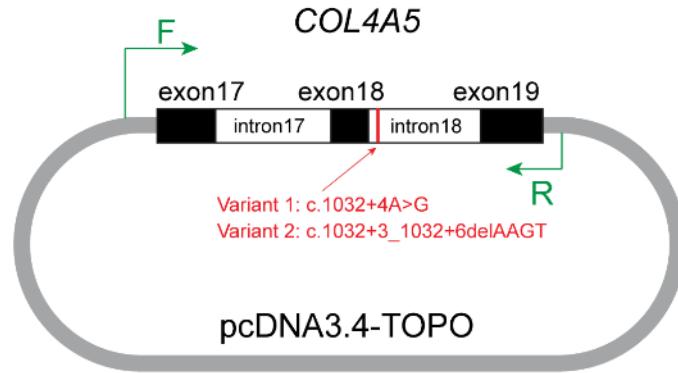
COL4A5
c.1032+3_1032+6delAAGT



COL4A5
c.1032+4A>G



Immunostaining of collagen 4



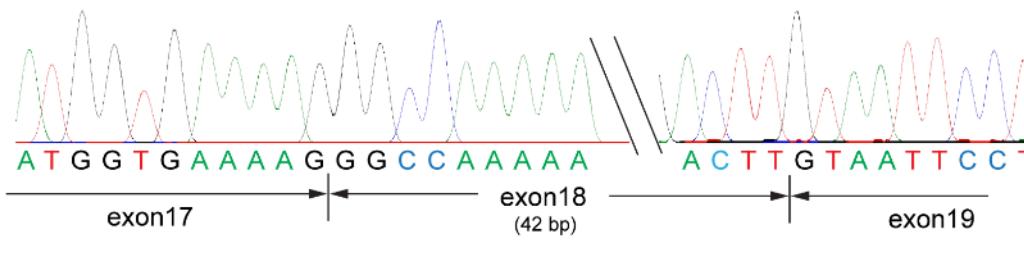
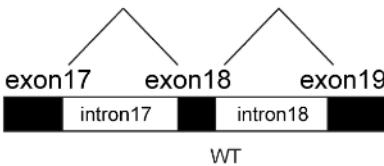
Zhang Yaochun



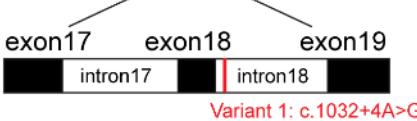
Tina Lim

Mini-Gene assay

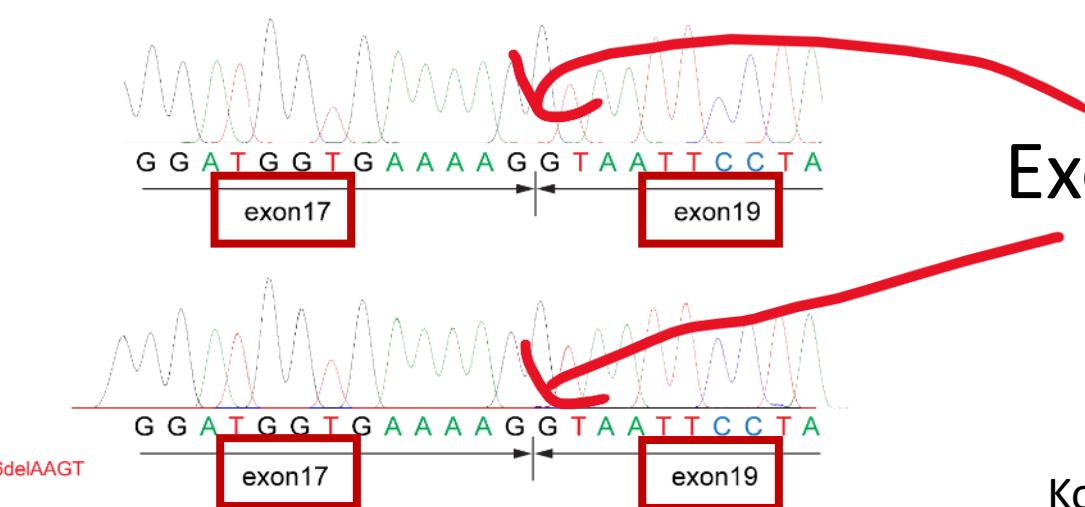
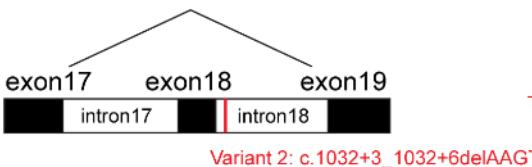
Wild type



Variant 1

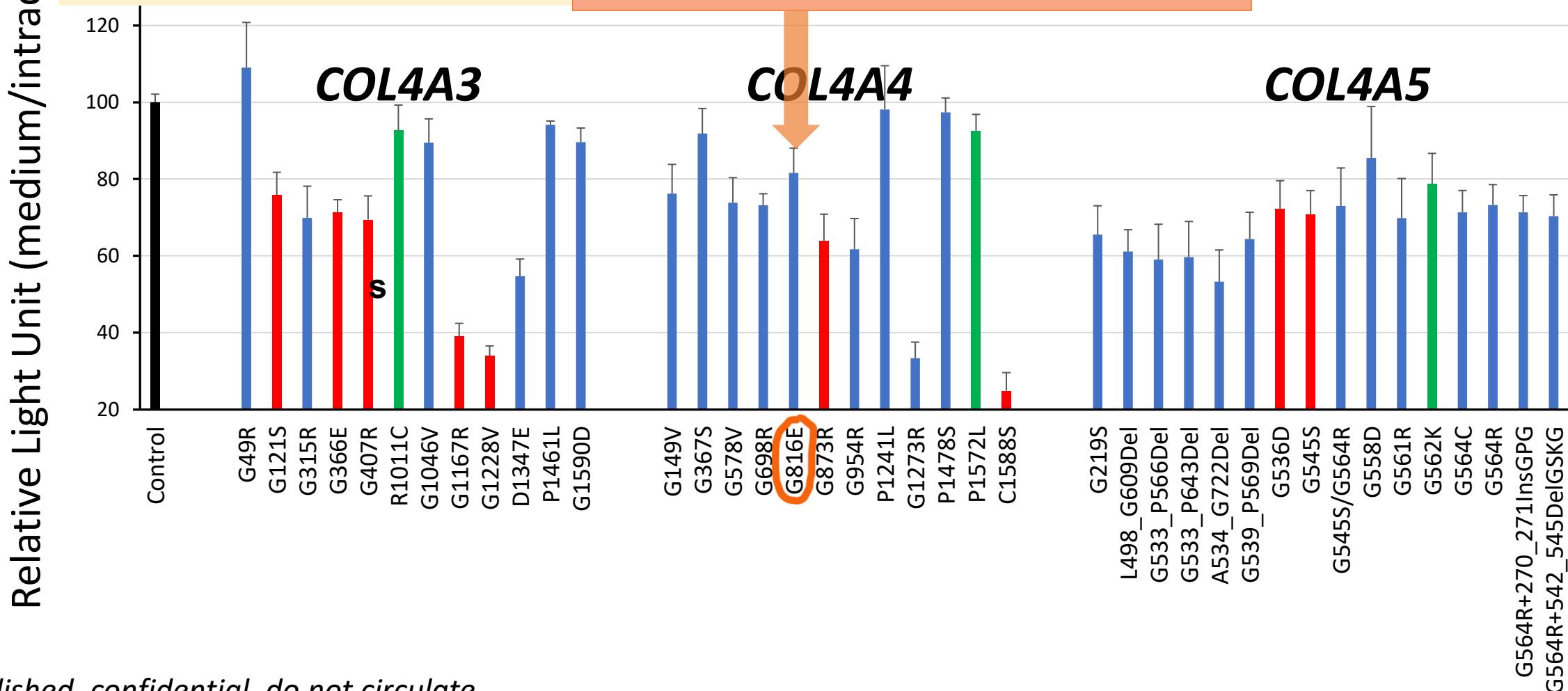


Variant 2



Split-luciferase assay

■ (Likely) benign
■ (Likely) pathogenic
■ Variant of uncertain significance



COL4A4, c.2447G>A p.G816E
GnomAD: 0.0334% in East Asians
SG10K All: MAF 0.0736%
SG10K Chinese: MAF 0.1221%



Zhang Yaochun



Tina Lim

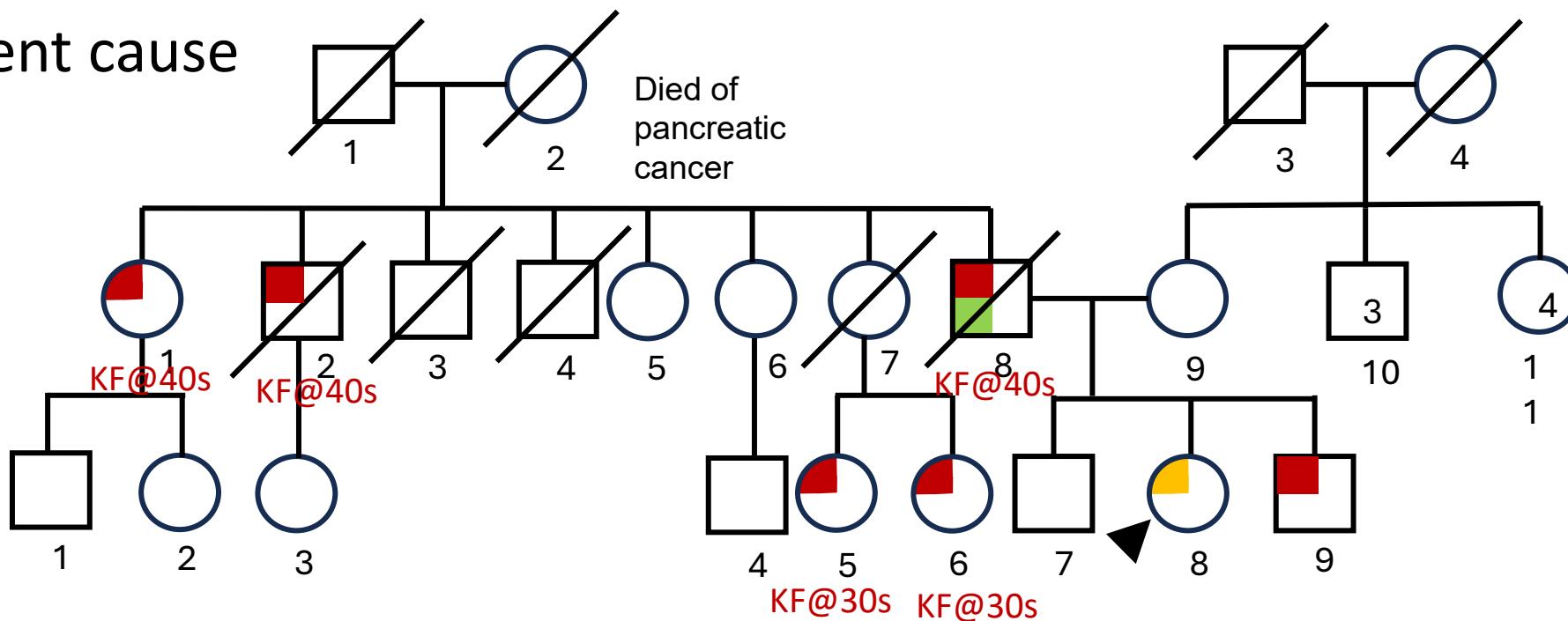
Outline: Interpreting genetic reports

- Basic 101: What nephrologists should know
- Understanding the limitations and caveats

31 year old Chinese lady

- Incidental CKD Stage G3A1 at age 29
- Ultrasound: Increased bilateral kidney echogenicity
- Bland urine sediments
- No apparent cause

Exome: no pathogenic variants
Biopsy: Tubulointerstitial nephritis



MUC1 (Mucin-1) gene testing

Specific Cytosine duplication

Diagnosed:

Mucin-1 related autosomal dominant tubulointerstitial kidney disease (ADTKD)

Variable number tandem repeat (VNTR)

Confidential, unpublished, do not circulate

3 year old Chinese girl

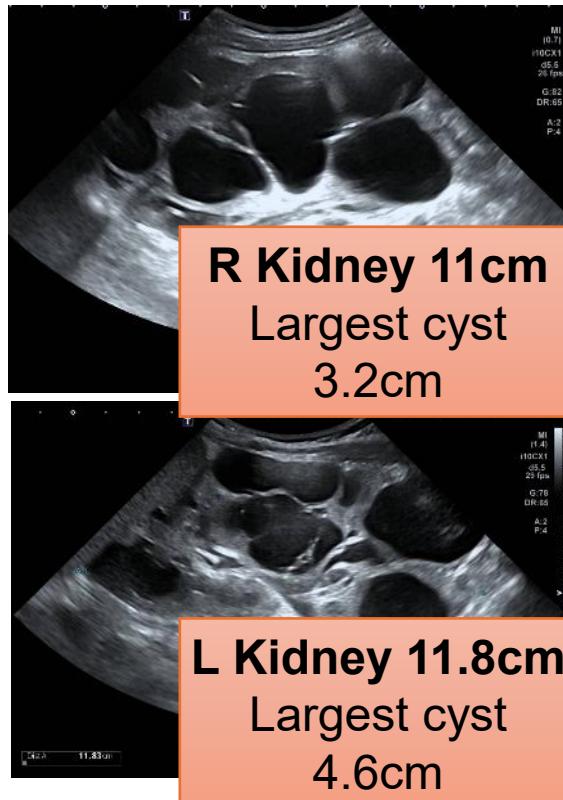
- Known **tuberous sclerosis** (classic skin features)
- Renal cysts** since 1.5 years old
- CKD G1A1
- No family history

Cystic gene panel

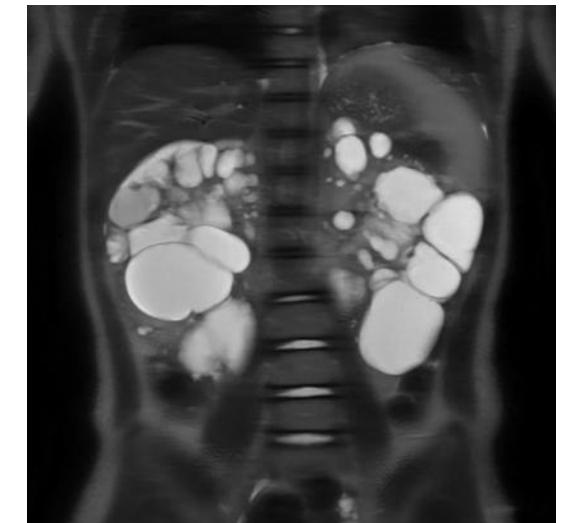
TSC2 gene (autosomal dominant):
Pathogenic mutation **5'UTR_3'UTRdel**

PKD1?

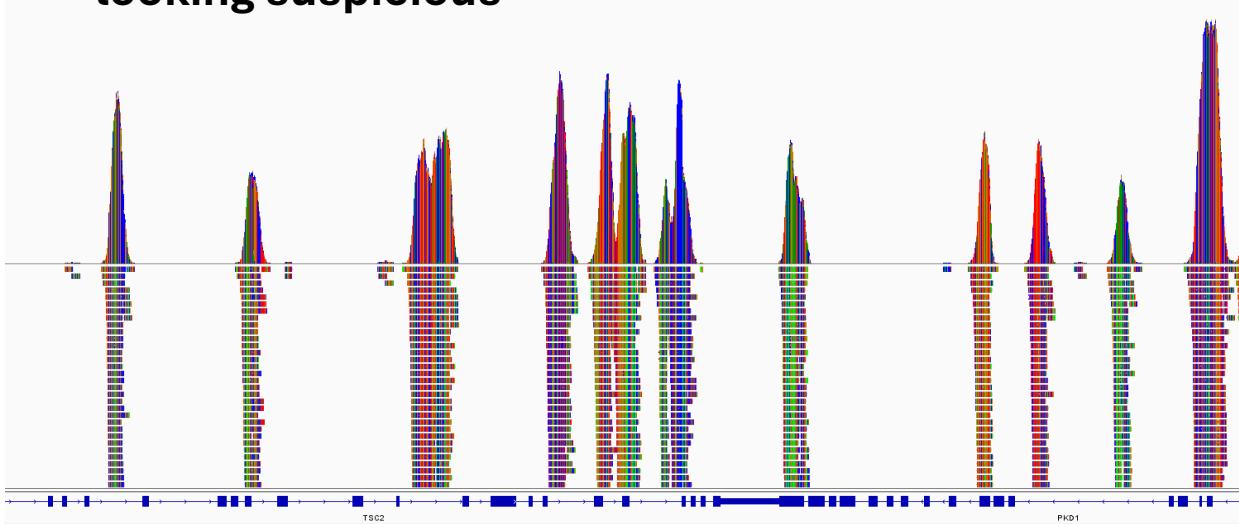
Ultrasound



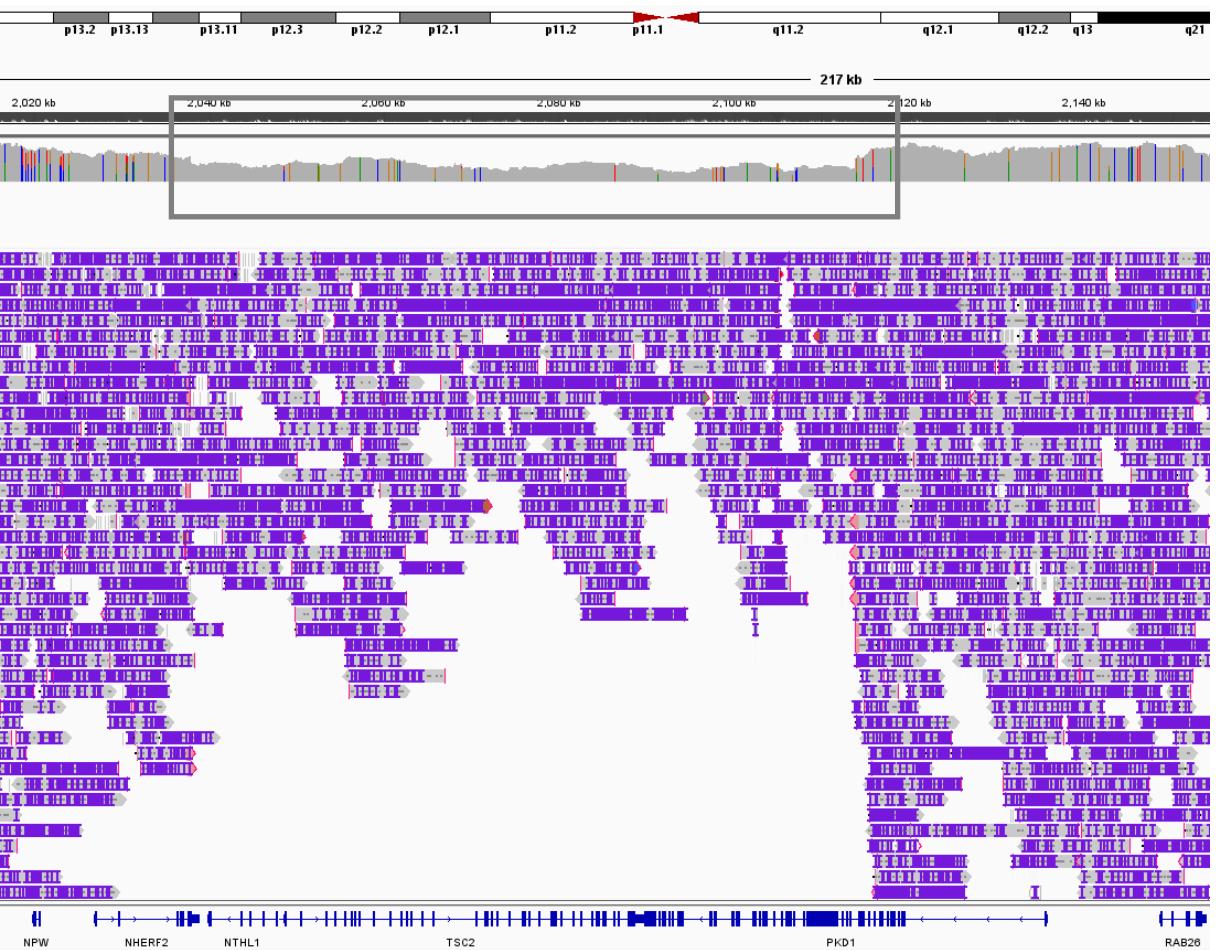
MRI



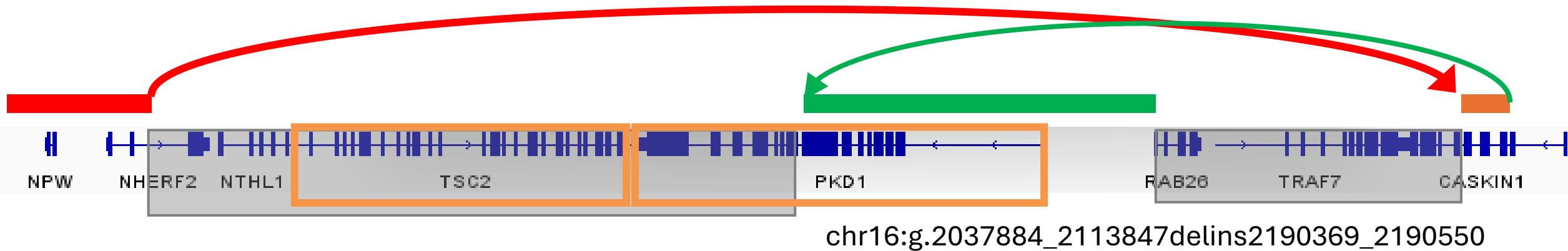
Exome short read sequencing: looking suspicious



Long read sequencing



Long read sequencing



Complete deletion: *TSC2*, *NTHL1*, *TRAF7*

Partial deletion: *PKD1*, *NHERF2*

Partial duplication: *CASKIN1*

Homologous recombination

**Diagnosed:
*TSC2-PKD1 contiguous tuberous sclerosis-polycystic kidney disease syndrome***

Which type of genetic tests should be ordered in patients with kidney diseases?

PHENOTYPE

CLEAR

?

UNCLEAR
OR CKDX

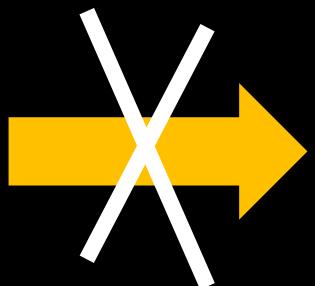
Panels
~50-150 genes

Kidney panel
~400-500 genes

Exome or Genome
~22,000 genes

Same diagnostic
yield as exome

Negative
genetic test



No genetic
disease

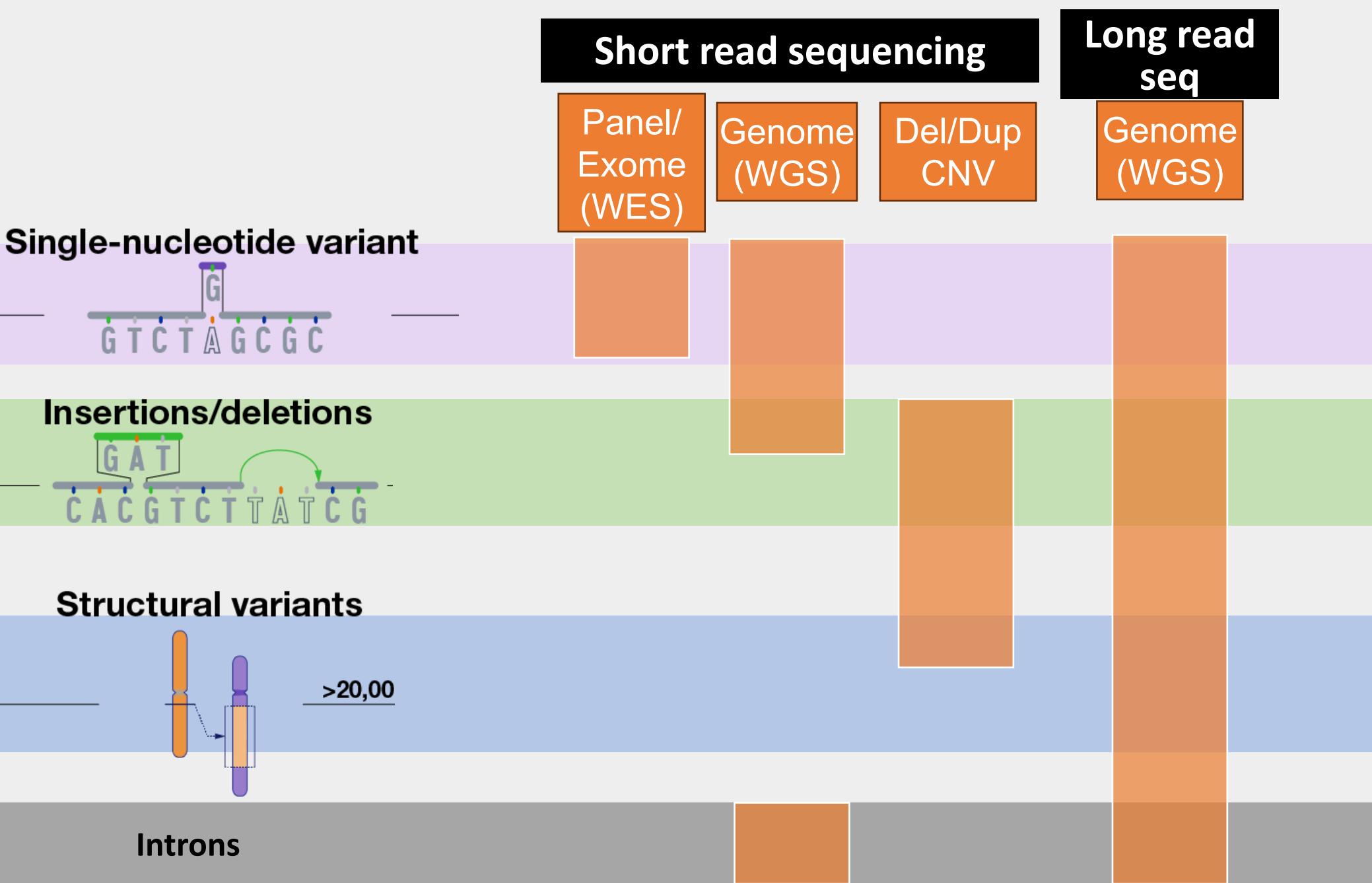
Variant problem

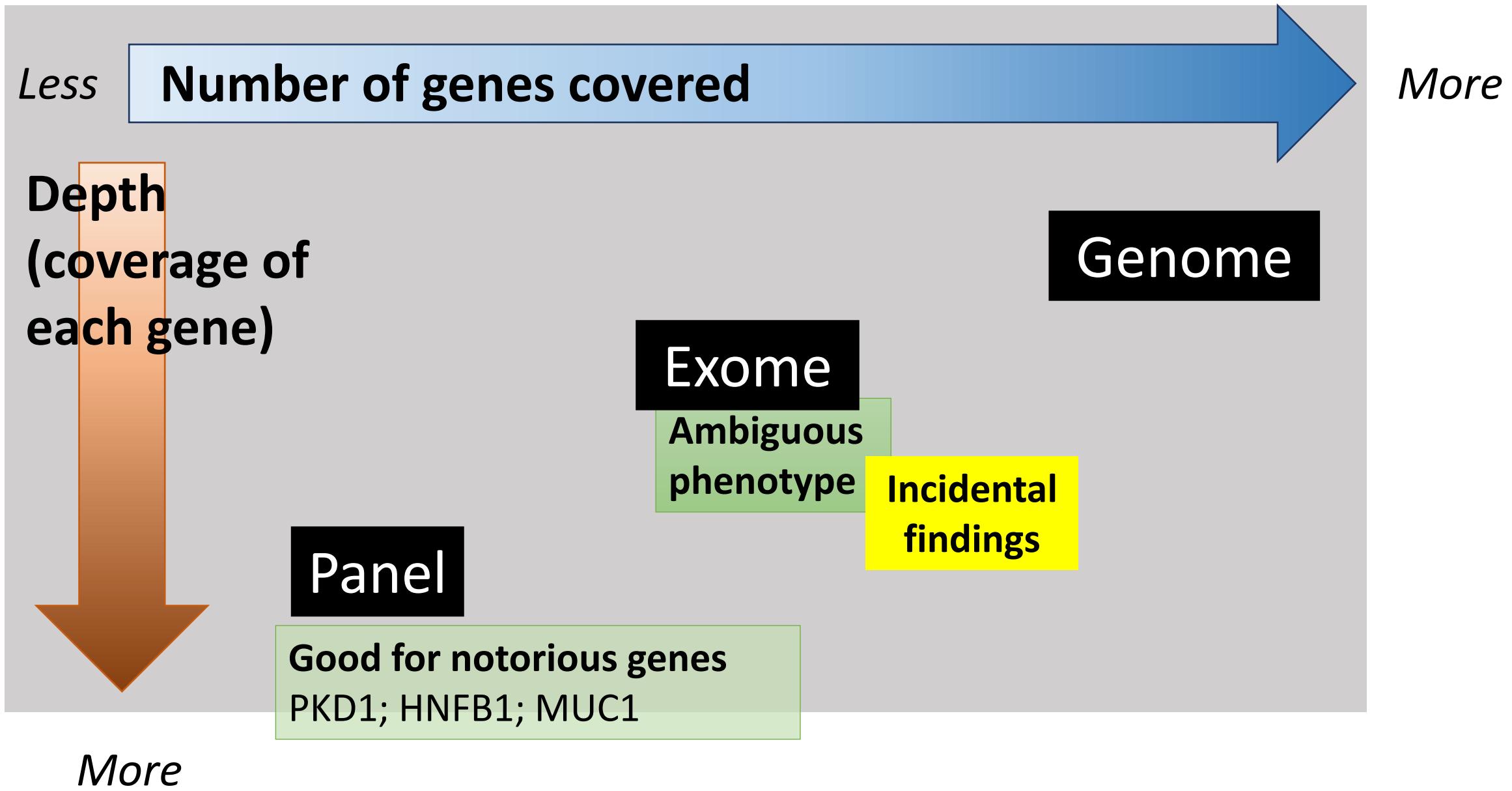
Variant of uncertain significance

Non-coding variants (e.g. splice)

Structural variant (e.g. large deletion / duplication)

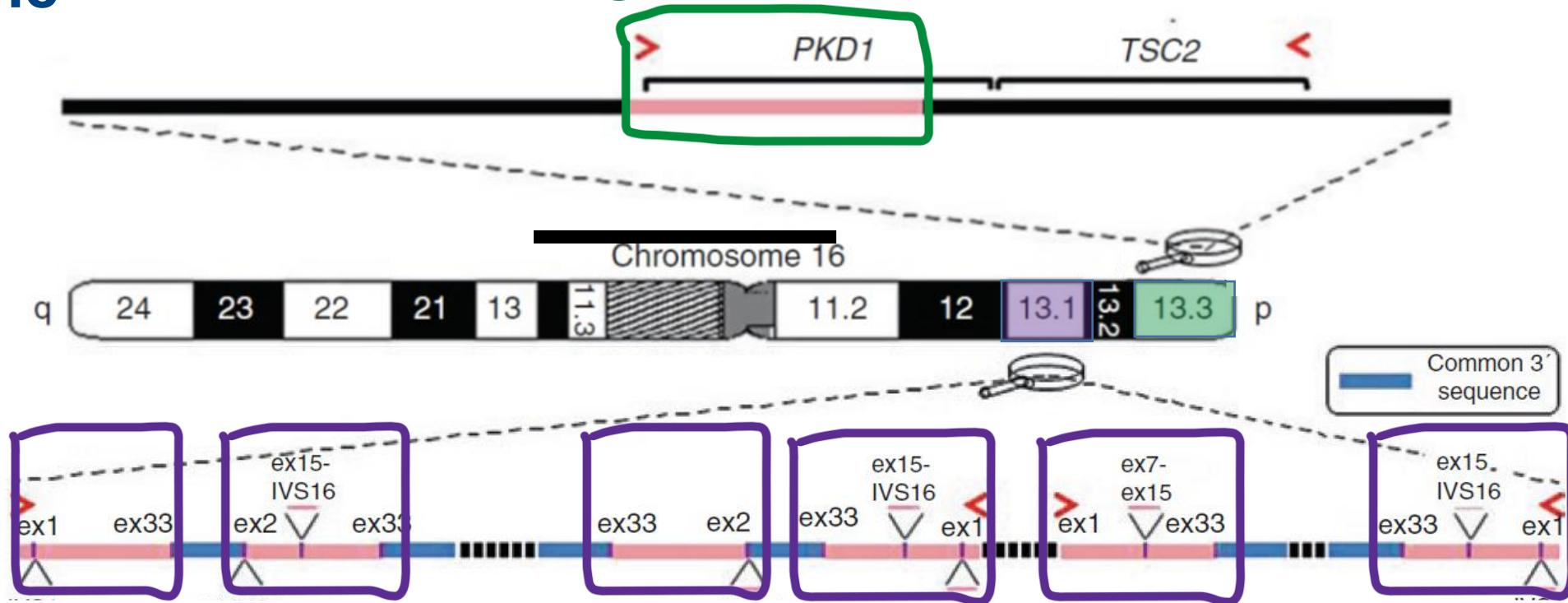
Gene not in the panel or not yet associated with diseases





PKD1 gene

Real gene, exons 1-46



6 pseudogenes exons 1-33
98% homology as the real gene

Is it in the real gene or the pseudogenes?

Whole exome sequencing: usually bad PKD1 coverage



TAKE HOME MESSAGES



1. Check the adequacy of a genetic report
 - Type of test, gene list, gene coverage
2. Check consistency in gene-phenotype and mode of inheritance

3. Resolution of variants of uncertain significance requires multi-disciplinary efforts and is iterative
4. Limitations of genetic tests:
 - **Genes:** Missing, challenging gene regions (pseudogenes, multiple repeats)
 - **Variants:** Non-coding, large deletions / duplications

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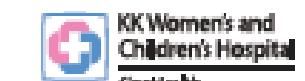
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